SETA BioMedicals Fluorescent Tools for BioMedical Applications

General Protein Labeling Procedures

A stock solution of 1 mg of the NHS-activated dye in 100 - 200 μ L of anhydrous dimethyl formamide (DMF) or dimethyl sulfoxide (DMSO) is prepared. Then aliquots of 5, 10, 25, and 50 μ L of this dye stock solution are added slowly to a solution of 5 mg IgG dissolved in 1 ml of a 50 - 100 mM bicarbonate buffer (pH 8.0 - 9.0). This mixture is allowed to stir for an additional 1 - 3 h at 25 °C. However, in most cases the labeling reaction will be completed within 5 - 10 minutes. To increase the degree of labeling a higher starting ratio of NHS-ester vs. protein should be used.

As the number of amino-groups in every protein is different it is important to adjust the D/P starting ratios in order to find the appropriate degree of labeling (DOL). The protein solution used for labeling should be free of amines and a TRIS buffer is therefore not suitable as a labeling buffer for NHS-esters. Antibodies stored in buffers containing amines are to be dialyzed against the labeling buffer (phosphate-buffered saline (PBS), or sodium bicarbonate).

Purification of Dye-Protein Conjugates

Separation of the dye-protein conjugate from non-conjugated dye is achieved using gel permeation chromatography on a 1.5 x 25 cm column (stationary phase: Sephadex G-25; eluent: 67 mM PB, pH 7.4). The fraction with the shortest retention time containing the colored dye-protein conjugate is collected. This first colored band will be the desired conjugate. The second, slower moving band in general contains the unlabeled free dye (hydrolyzed NHS-ester).

Determination of Dye-to-Protein Ratio (D/P)

This procedure including the x-factor values are provided in a separate technical note on our website.

Storage of Dye-Protein Conjugates

Dye-protein conjugates are to be stored under similar conditions as used for the unlabeled protein. Typical storage temperatures are 4℃ and sodium azide can be added to avoid bacterial growth. For long-term storage, prepare smaller aliquots and freeze. Avoid repeated freezing and thawing. Protect from light.